### **Review Article**

# The aetiology of cardiovascular disease: a role for mitochondrial DNA?

Marianne Venter, Francois H van der Westhuizen, Joanna L Elson

#### Abstract

Cardiovascular disease (CVD) is a world-wide cause of mortality in humans and its incidence is on the rise in Africa. In this review, we discuss the putative role of mitochondrial dysfunction in the aetiology of CVD and consequently identify mitochondrial DNA (mtDNA) variation as a viable genetic risk factor to be considered. We then describe the contribution and pitfalls of several current approaches used when investigating mtDNA in relation to complex disease. We also propose an alternative approach, the adjusted mutational load hypothesis, which would have greater statistical power with cohorts of moderate size, and is less likely to be affected by population stratification. We therefore address some of the shortcomings of the current haplogroup association approach. Finally, we discuss the unique challenges faced by studies done on African populations, and recommend the most viable methods to use when investigating mtDNA variation in CVD and other common complex disease.

Keywords: mitochondrial DNA, cardiovascular disease, MutPred, mutational load, African

Submitted 7/11/16, accepted 31/7/17 Published online 24/8/17 *Cardiovasc J Afr* 2017; **29**: 122–132

www.cvja.co.za

DOI: 10.5830/CVJA-2017-037

#### **Mitochondrial DNA**

Cardiovascular disease (CVD) remains the main noncommunicable cause of morbidity and mortality in humans.<sup>1</sup> While environmental factors and lifestyle choices play a major role in CVD, it is also recognised that genetic factors contribute significantly to the aetiology thereof. In this regard, several studies, most recently genome-wide association studies (GWAS),

Human Metabolomics, North-West University, Potchefstroom, South Africa Marianne Venter, PhD, 20196946@nwu.ac.za Francois H van der Westhuizen, PhD Joanna L Elson, PhD

Institute of Genetic Medicine, Newcastle University, United Kingdom

Joanna L Elson, PhD

have contributed to identifying genetic loci involved in CVDs, and their association with behavioural and biological risk factors.<sup>2-7</sup>

Despite the numerous nuclear DNA (nDNA) variants identified, only a small portion of the heredity of CVDs can thus far be accounted for by variants discovered with GWAS studies.<sup>8</sup> For instance, the 46 loci identified for coronary artery disease (CAD) only account for about six to 13% of CAD hereditability.<sup>9-11</sup>

The mitochondrion is the only other source of DNA apart from the nucleus. Mitochondrial DNA (mtDNA) encodes for 22 tRNAs, two rRNAs and 13 polypeptides thought important in the catalytic cores of complexes I, III, IV and V of the oxidative

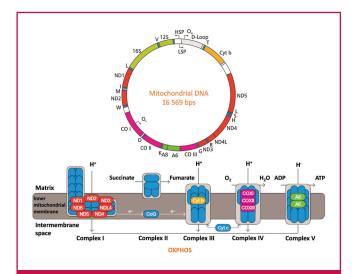


Fig. 1. mtDNA encodes for 22 tRNA and two rRNA molecules. as well as 13 polypeptide sub-units of the OXPHOS enzyme complexes, as indicated by colour. Enzyme complexes I-IV are involved in a series of redox reactions, which transfer electrons from carriers nicotinamide adenine dinucleotide (NADH) and flavin adenine dinucleotide (FADH<sub>2</sub>) to oxygen molecules. During these catalytically favourable reactions, H<sup>+</sup> ions are pumped from the mitochondrial matrix into the mitochondrial intermembrane space to create a proton-motor force across the inner mitochondrial membrane. This force is used by complex V to catalyse the phosphorylation of adenosine diphosphate (ADP) to adenosine triphosphate (ATP). Complex I: NADH dehydrogenase; complex II: succinate dehydrogenase; complex III: cytochrome c reductase; complex IV: cytochrome c oxidase; complex V: ATP synthase.

phosphorylation (OXPHOS) system (Fig. 1). In humans, mtDNA contains 16 569 bps and is double stranded.<sup>12</sup> Depending on the energy needs of a specific tissue, each cell can contain hundreds to thousands of copies of mtDNA.<sup>13</sup> mtDNA is maternally inherited and has a much higher mutation rate than nDNA, possibly 10 to 17 times higher.<sup>14</sup> Maternal inheritance results in a lack of bi-parental recombination and therefore the evolution of mtDNA is defined by the emergence of distinct lineages called haplogroups.

Multi-copy makes possible a condition called heteroplasmy, where more than one genotype is present in the same cell/ tissue/organism; homoplasmy then, is where all mtDNA copies carry the same allele. Notably, mtDNA is largely overlooked in GWASs, and could possibly contribute to the missing heredity of CVDs. Next we will consider two main arguments on the possible role of mtDNA variants in CVDs.

### Mitochondrial dysfunction and mtDNA damage in vascular health

When considering mtDNA as a possible contributor to the aetiology of CVD, it should also be considered from a biological perspective. Much investigation has been conducted in an attempt to elucidate the risk factors and physiological mechanisms involved in the development of CVDs, such as sub-clinical atherosclerosis, hypertension, cardiomyopathy and type 2 diabetes.<sup>15-20</sup>

An important common feature in all these conditions is inflammation in some form or another (Fig. 2). This inflammatory state is thought to be caused by oxidative stress, due to excessive levels of reactive oxygen species (ROS). ROS can be produced in several pathways, including by enzymes such as NADPH oxidase, nitric oxide synthase, and enzyme complexes of the electron transport chain (ETC).<sup>21</sup>

The general mechanism of ROS involvement in CVDs is ascribed to oxidative effects. For example, ROS contributes to atherosclerotic lesion formation by oxidising lipids, promoting vessel wall uptake of inflammatory cells, and enhancing proliferation and hypertrophy of vascular smooth muscle cells (VSMC).<sup>21</sup>

Several studies have shown increased levels of ROS in hypertensive humans and rats.<sup>16,22,23</sup> In cultured VSMCs for example, ROS has been shown to cause changes in cellular signalling pathways, favouring vasoconstriction.<sup>15</sup> A mechanism for this could be that ROS reduces nitric oxide (NO) bioavailability via quenching, impairing endotheliummediated vasodilation.<sup>21,22,24</sup> However, ROS along with other factors of a dysfunctional mitochondrial energy metabolism (e.g. nucleotides, Ca<sup>2+</sup>) also act as effectors of retrograde signalling and the so-called cell danger response.<sup>25,27</sup>

Mitochondria are considered the major producers of ROS within the cell. In a recent article, Lopez-Armada *et al.*<sup>18</sup> reviewed the role of mitochondrial dysfunction in the inflammatory response and consequently in the pathology of various diseases, including CVDs. The authors described how mitochondrial dysfunction may modulate inflammatory processes by activating redox-sensitive inflammatory pathways and the NLRP3 inflammasome. In the vasculature, these alterations lead to disturbed endothelial homeostasis, which has been implicated in the pathology of CVDs, such as atherosclerosis.<sup>18</sup> Indeed,

some improvements in disease presentation of hypertension and diabetes have been observed in studies where chronic antioxidant treatment is applied.<sup>18,28,29</sup>

Another mechanism by which inflammation might be altered by mitochondrial dysfunction is through the resultant release of mtDNA into the cytosol and circulation. Because mtDNA is similar to bacterial DNA and not methylated,<sup>30</sup> released mtDNA molecules are thought to induce an inflammatory state, which contributes to atherosclerosis and other inflammatory diseases.<sup>31-35</sup>

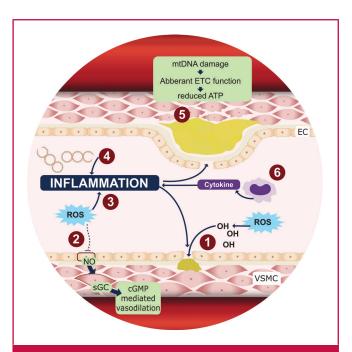


Fig. 2. Mitochondrial dysfunction and mtDNA damage affect vascular health in several ways. (1) ROS aids in lesion formation by oxidising lipids, increasing the uptake of inflammatory cells into the vascular wall and enhancing proliferation and hypertrophy in VSMC. (2) During endothelium-dependant vasodilation, EC-released NO activates sGC in VSMC to produce cGMP, signalling a vasodilation response. ROS inhibits this mechanism by quenching bioavailable NO molecules. (3) Endothelial homeostasis is disturbed and plaque formation promoted when mitochondrial dysfunction leads to ROS formation and activates redox-sensitive inflammatory pathways. (4) Circulating cell-free mtDNA is similar in structure to bacterial DNA and invokes an inflammatory response, contributing to atherosclerosis. (5) Independent from ROS formation, mtDNA damage leads to aberrant ETC function and reduced ATP levels in VSMC. When cell viability is compromised, apoptosis of VSMC occurs, accelerating plaque growth and decreasing plaque integrity. (6) Through the same mechanisms, apoptosis of monocytes occurs, releasing inflammatory cytokines, contributing to inflammation and consequently, increasing plaque formation and vulnerability. ATP: adenosine triphosphate; cGMP: cyclic guanosine monophosphate; EC: endothelial cell; ETC: electron transport chain; NO: nitric oxide; ROS: reactive oxygen species; sGC: soluble guanylyl cyclase; VSMC: vascular smooth muscle cells.

mtDNA damage has also been shown to promote atherosclerosis directly, in the absence of oxidative stress. In a study by Yu *et al.*,<sup>36</sup> VSMCs showed increased apoptosis and decreased proliferation in a proof-reading deficient PolG<sup>4/</sup> ApoE<sup>4/</sup> mouse model. Increased secretion of pro-inflammatory factors, tumour necrosis factor- $\alpha$  and interleukin-1 $\beta$  were also reported and implicated in mtDNA release into the cytosol and subsequent activation of the inflammasome.

The authors went on to test the applicability of their findings in humans and concluded that an alternative mechanism for mtDNA defects mediate atherosclerosis development, independent of ROS; mtDNA defects lead to aberrant ECT function and consequently reduce ATP content in VSMCs, which then promote apoptosis and inhibit cell proliferation, leading to increased atherosclerosis and risk of plaque rupture.<sup>36,37</sup> Plaque vulnerability is further promoted by mtDNA defects via monocyte cell death and the resultant increased release of inflammatory cytokine.<sup>38</sup> From these studies, it can be seen that mitochondrial dysfunction, possibly as a result of mtDNA variants or damage, can directly be implicated in mechanisms that encumber vascular health.

#### mtDNA point mutations and cardiac involvement

Clinically proven mtDNA mutations are also an important cause of inherited disease.<sup>39</sup> To date, more than 250 deleterious point mutations and deletions of the mitochondrial genome have been clinically proven to be associated with certain disease phenotypes (www.mitomap.org). In several of these diseases, cardiovascular symptoms are an important part of the aetiology.

Due to the very high levels of mtDNA population variation seen, both within and between human populations, the identification of mutations causing clinically manifesting disease proves to be a challenge, despite the small size of the mitochondrial chromosome. Initially, DiMauro and Schon had set specific criteria for defining the pathogenicity of mtDNA mutations.<sup>40</sup> The list has subsequently been updated to include important methods such as functional testing and single-fibre analysis, which can more specifically link genotype to phenotype.<sup>41,42</sup>

Notably, a pathogenicity scoring system for mitochondrial tRNAs was devised by McFarland *et al.*<sup>41</sup> and further refined by Yarham *et al.*<sup>43</sup> Mitchell *et al.*<sup>44</sup> also devised a pathogenicity scoring system using variants in complex I mtDNA genes, but this can be applied to any structural mtDNA mutation. A list of these criteria is given in Table 1.

It should be noted that there are mtDNA mutations that are exceptions to all the 'rules' in Table 1, and this was a critical motivation for algorithms or clinical scoring systems to help weigh the evidence that is presented for each mutation.<sup>43,44</sup> For a clinically proven mutation to manifest as a diseased phenotype, as in the case of primary mitochondrial disorders, the allele frequency (heteroplasmy) needs to exceed a certain threshold, usually above 60%, referred to as the phenotypical threshold effect.<sup>45</sup>

The biochemical threshold effect then refers to the ability of the oxidative phosphorylation (OXPHOS) system to resist the metabolic expression of deficiencies therein.<sup>45,46</sup> These deficiencies may be caused by various factors involved in the expression and regulation of the OXHPOS complexes. There are many complexities to the expression of mtDNA mutations; a classic example is the mitochondrial tRNA mutation m.3243A>G, the most common of the mtDNA mutations causing mitochondrial disease. The m.3243A>G mutation can result in a vast array of clinical phenotypes affecting multiple systems within the body, causing two distinct clinical syndromes: maternally inherited diabetes and deafness (MIDD), and mitochondrial encephalomyopathy, lactic acidosis, and stroke-like episode (MELAS) syndrome in severe cases. Furthermore, the age of onset of m.3243A>G-associated phenotypes spans more than 50 years. The impact of several confounding factors, including heteroplasmy levels, remains unclear.<sup>47</sup>

Another group of well-studied mutations are those that cause the disease Leber's hereditary optic neuropathy (LHON). In contrast to the m.3243A>G mutation, LHON has a tissue-specific phenotype manifesting as bi-lateral blindness. Several mtDNA mutations have been implicated in LHON, while three of these mutations, namely m.3460G>A, m.11778G>A and m.14484T>C located in subunits ND1, ND4 and ND6 of complex I, respectively, account for 90 to 95% of cases.<sup>48</sup> Unusually, these mutations can be detected as homoplasmic variants without exerting a phenotype. Rather, disease penetrance is significantly influenced by confounding factors such as gender and environment (clinical penetrance is increased to 93% in smoking men),<sup>49</sup> and mtDNA haplogroup background (haplogroup J, K and M7 increase risk of clinical penetrance).<sup>50,51</sup>

The heart has especially high energy needs and relies heavily on OXPHOS-derived ATP, such that one-third of cardiomyocyte volume consists of mitochondria.<sup>52</sup> Not surprisingly then, the myocardium is frequently affected in primary mitochondrial disorders.<sup>53</sup> In a retrospective review study by Yaplito-Lee *et al.*,<sup>54</sup> 33% of paediatric patients with definitive OXPHOS disorders had cardiac manifestations. Several mtDNA mutations (Fig. 3, Table 2 [online]) have also been shown to exhibit cardiac involvement, either as part of a multi-system syndrome (most frequently seen in MELAS), or as isolated occurrences, such as in the absence of associated CVDs or risk factors thereof.<sup>53,55,66</sup>

Hypertrophic cardiomyopathy (hCM) and pulmonary artery hypertension (PAH) are the two phenotypes most commonly seen as isolated cardiac manifestations of primary mitochondrial

#### Table 1. Criteria for defining the pathogenicity of mtDNA mutations

- Criteria for pathogenicity of mtDNA mutations include
- The mutation must be present only in patients and not in controls
- The mutation must be present in varied mitochondrial genetic backgrounds
- The mutation must be the best mtDNA candidate variant to be pathogenic
- The mutation must affect functionally important domains
- Transfer of the mutated mtDNA to another cell line must be accompanied by transfer of the cellular or molecular defect
- The mutation must not be a recognised, non-pathogenic, single-nucleotide polymorphism
- The mutation must alter an area that is known to be highly conserved throughout evolution
- The mutation must occur at varying levels within the cells (i.e. must be heteroplasmic)
- A larger proportion of mutant mtDNA must correspond to a more severe phenotype
- Single-fibre polymerase chain reaction must be performed by comparing normal and abnormal fibres from muscle
- The secondary structure of the tRNA molecule must also be taken into account when determining mt-tRNA mutation pathogenicity

These criteria need to be met in order for a mtDNA mutation to be classified as 'disease causing' for either structural or mt-tRNA mutations.<sup>40-43</sup>

disorders.<sup>53</sup> If clinically proven mtDNA mutations can directly lead to cardiac dysfunction, is it plausible to think that other mtDNA variants, such as population variants of mildly deleterious effect, might also lead to or alter severity/penetrance of complex cardiovascular disease phenotypes.

From the substantial supportive evidence of mitochondrial involvement in cardiovascular disease, it is therefore evident that genetic investigations on the aetiology of CVD should include consideration of mtDNA variations. In the following sections, we present a number of approaches (plus findings from such investigations) on how mtDNA variation is investigated/ associates in/with disease, with a specific focus on the approaches more likely to show its putative contribution to the risk of CVD development.

## Current approaches used for investigating mtDNA involvement in disease

#### Mitochondrial DNA copy number

mtDNA copy number can be used as an indicative marker of mitochondrial biogenesis, which is thought to increase in response to increased energy demands, such as exercise, but also as a compensatory method for mitochondrial dysfunction.<sup>89</sup> On the other hand, mtDNA copy number has been shown to decrease with aging,<sup>90</sup> and has been significantly correlated

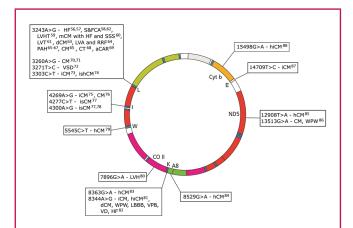


Fig. 3. mtDNA morbidity map indicating clinically proven mtDNA mutations that present with syndromic or isolated cardiac involvement. aCAR: abnormal cardiac autonomic regulation; CM: cardiomyopathy; hCM: hypertrophic cardiomyopathy; dCM: dilated cardiomyopathy; HF: heart failure; hiCM: histiocytoid cardiomyopathy; iCM: infantile cardiomyopathy; ishCM: isolated hypertrophic cardiomyopathy; LBBB: left bundle branch block; LVA: left ventricle abnormalities; LVH: left ventricular hypertrophy; LVHT: left ventricular hyper-trabeculation/ non-compaction; mCM: mitochondrial cardiomyopathy; PAH: pulmonary artery hypertension; RRF: ragged red fibres; S&FCA: structural and functional cardiac abnormality; SSS: sick sinus syndrome; VD: ventricular dysfunction; VPB: ventricular premature beats; VSD: ventricular septal defect; WPW: Wolff-Parkinson-White syndrome. See Table 2 for a detailed list of mutations, phenotype, references and pathogenicity scores, as described in Mitchell et al.44 and Yarham et al.43

with late-onset diseases, such as Parkinson's disease.<sup>91,92</sup> As mentioned above, cell-free circulating mtDNA may also act as an inflammatory agent that contributes to CVDs.<sup>33</sup>

Altered mtDNA copy number measured in peripheral blood cells have been shown to be associated with different complications of diabetes (diabetic retinopathy and diabetic nephropathy).<sup>93,94</sup> Also, an association between telomere length and mtDNA copy number suggests a co-regulatory mechanism for these two parameters, both of which are implicated in aging.<sup>95</sup> mtDNA depletion and impaired mitochondrial biogenesis have been shown to be a constant factor in the early stages of heart failure<sup>96,97</sup> and other diseases thought to be related to aberrant ROS production.<sup>98</sup>

While the exact mechanisms behind mtDNA content regulation are still unclear, it seems changes in either direction can be causative or indicative of disease.<sup>99</sup> Measurement of mtDNA copy number can be done accurately by real-time PCR methods, making this a useful approach for investigating the role of mitochondrial metabolism in disease phenotypes.

#### **Common mtDNA population variants**

mtDNA variants accumulated over time differ between population groups that have been separated for several thousand years. Consequently, distinct lineages (mtDNA haplogroups) can be drawn according to these sets of unique changes in mtDNA, referred to as common population variants. The full human mtDNA phylogeny can be accessed at www.phylotree.org.<sup>100</sup> Much of the variation seen in modern humans is to be found in the African haplogroups L0 to L6, but this variation has not been as fully described as the variation on other continents. European (e.g. I, J, K, H, T, U, V, W, X) and Asian (e.g. A, B, C, D, F, G) haplogroups fall within super haplogroups M and N, which in turn fall within L3.

mtDNA haplogroup association studies aim therefore to associate these common mtDNA population variants with risk for various complex diseases, such as diabetes, hypertension or Parkinson's disease.<sup>101</sup> mtDNA background has been shown to correlate with the severity of cardiomyopathy caused by nDNAencoded mitochondrial protein mutations,<sup>102</sup> and increases the penetrance of LHON-causing pathogenic mutations.<sup>50,51</sup>

It has been proposed that mtDNA population variants could contribute to the adaptability of population groups to their environment by altering mitochondrial enzyme function.<sup>103,104</sup> By analysing non-synonymous variants in 104 complete mtDNA sequences from across the globe, Mishmar et al.<sup>103</sup> found that the ATP6 and cytochrome b genes were particularly variable in arctic and temperate zones, respectively, leading them to believe that positive selection had taken place. Stressors, such as sudden changes in environment, could then influence the degree of disease susceptibility of these environmentally adapted population groups.<sup>105</sup> However, this hypothesis was contested by others who have shown that there are significant differences in the same measure in haplogroups from the same environment.<sup>106,107</sup> Additionally, Amo and Brand<sup>108</sup> put forward evidence to suggest that certain bioenergetic parameters did not significantly differ between mitochondria from arctic versus tropical haplogroups.

In contrast to the action of positive selection, the action of negative or purifying selection on mtDNA has been established for almost a decade.<sup>107,109</sup> One important point to consider is that

positive or directional selection could not have acted identically on all lineages, and therefore would result in a different rate of accumulation of variants on haplogroup lineages, thus affecting our ability to time divergence events by counting the mutational events between lineages. On the other hand, it is possible that negative or purifying selection could act evenly across lineages and not impact on our use of mtDNA as a molecular clock; the reliability of mtDNA as a molecular clock has been widely discussed.<sup>110</sup>

Because of the central role that mitochondria play in cell signalling and apoptosis, mitochondria have been implicated in several age-related diseases, including Parkinson's disease, Alzheimer's disease, multiple sclerosis and psoriasis.<sup>101,111,112</sup> CVDs are also classified as late-onset diseases and mitochondria have also been implicated in CVDs. Consequently, haplogroup association studies on CVD phenotypes are plentiful, but, as will be revealed, also prone to pitfalls.

Crispim *et al.*<sup>113</sup> reported an association of European haplogroup cluster J/T with insulin resistance and type 2 diabetes in a Caucasian-Brazilian cohort. On the other hand, Li *et al.*<sup>114</sup> found no association between mtDNA variation and risk for developing diabetes, while Chinnery *et al.*<sup>115</sup> found no association with type 2 diabetes and major European haplogroups in a large study using 897 cases and 1 010 controls. Rather, Achilli *et al.*<sup>116</sup> found that the risk for developing specific types of diabetes complications (disease outcome) was significantly associated with different mitochondrial haplogroups.

Several mtDNA population variants in cytochrome c oxidase and NADH dehydrogenase subunit genes have been associated with body mass index (BMI) in adults.<sup>117</sup> In a very large study using a second cohort, Chinnery *et al.*<sup>118</sup> found no significant associations between mtDNA haplogroups and ischaemic heart disease, hypertension, diabetes or the metabolic syndrome, but did find a significant association of sub-haplogroup K with risk of cerebral ischaemic vascular effects.

Therefore, while some studies investigating phenotypes included in CVDs have reported results that support a role for mtDNA in CVD,<sup>116,117,119,120</sup> there are also conflicting reports.<sup>115,118,121</sup> This is not only common in CVD-related literature, but all areas where haplogroup association studies have been applied. This is an indication of the many difficulties that need to be overcome when considering mtDNA variation in the context of disease.<sup>122</sup>

The unique way in which mtDNA is inherited (lack of bi-parental recombination), which results in the emergence of numerous unique haplogroups, contributes to the complexity encountered when investigating mtDNA involvement in disease. Non-biological factors such as differences in approach to statistical analysis;<sup>123</sup> difficulty in proper case and control matching; small effective population size, which results in a higher likelihood of population stratification; and insufficient cohort size,<sup>122</sup> further undermine the consistency of these studies.

Meta-analysis of data generated by several studies with overlapping phenotypes can be employed to overcome sample size difficulties, but these bring along challenges of their own, as independent studies have different goals/methods, and do not necessarily generate directly comparable datasets.<sup>101</sup> So, while haplogroup association studies might have fulfilled an important role in the ongoing pursuit of the involvement of mtDNA variation in disease, it is now well recognised that the field needs to consider alternative models.

#### **Rare mtDNA population variants**

It has been shown that negative or purifying selection plays a significant role in mtDNA evolution, with deleterious variants being removed from the population over time,<sup>107</sup> and that the power of selection has been equally effective in all human lineages.<sup>124</sup> Consequently, rare mtDNA population variants are more likely to be mildly deleterious than common variants, as selection has had less time to remove them from the populations. Indeed, rare mtDNA variants have been linked to changes in CVDs and risk factors.

In a study by Govindaraj *et al.*,<sup>120</sup> complete mtDNA analysis revealed 10 non-synonymous variants present in hypertrophic cardiomyopathy patients, but not present in controls or on databases. Seven of these variants were classified as likely 'pathogenic', using several online scoring tools such as PolyPhen-2, PMUT and PROVEAN, and were therefore thought to be involved in the development of cardiomyopathy. Rare variants m.5913G>A and m.3316G>A have both been suggested to be associated with increased fasting blood glucose levels, while m.5913G>A was shown to also be associated with increased blood pressure in a selected Framingham heart study subset, all of whom were of European descent.<sup>7</sup>

In addition, several rare mtDNA variants, such as m.3316G>A,<sup>7,125</sup> have been implicated in diabetes mellitus, of which an up-to-date list can be found on www.mitomap.org. Another possibility is that the effect of an accumulation of mildly deleterious variants may only become clinically significant once a population is challenged by a rapid change of confounding factors, such as diet or other environmental factors (toxins).<sup>126,127</sup>

In conclusion, several approaches are currently in use for investigating the role of mtDNA in common complex disease. mtDNA copy number is an emerging approach that might become more prevalent in studies concerning CVDs as well. In terms of mtDNA variants, rare population variants have been linked to several disease phenotypes, including CVD-related diseases such as cardiomyopathy and diabetes mellitus, and might be found to be associated with other CVDs or risk factors such as hypertension.

Rare population variants are more likely to be mildly deleterious,<sup>124</sup> but might not have a high enough impact on their own to alter disease onset; rather, these variants might be more likely to alter disease progression or outcomes. For common population variants, several haplogroup association studies have been done in CVDs, but have also been marred by the challenges these types of studies face.<sup>122</sup> It seems then that an alternative approach to investigating the role of mtDNA variation in disease is needed when investigating common complex disease.

### Alternative approach for investigating mtDNA involvement in disease: the adjusted mutational load hypothesis

An alternative approach, the mutational load hypothesis was put forward in Elson *et al.*<sup>111</sup> Mutational load refers to the synergistic effect of several changes in, for example, a specific gene or functionally related set of genes. It does not look for associations with a specific variant but rather a summative effect. While some mtDNA variants might be of little effect on their own, an increased mutational load might be associated with increased risk for a certain disease. mtDNA mutational loads can then

127

be adjusted to reflect the position within the phylogeny, since there are large differences in the average number of common population variants between haplogroups. This approach can also further be modified to, for example, exclude low-impact variants, highlighting the role of likely deleterious functional variants.

Determining the likely impact or pathogenicity of mtDNA variants can be achieved by using several computational pathogenicity-predicting methods.<sup>128</sup> An example of such a method is the MutPred system, which assigns a MutPred score to any protein-coding mtDNA variant, according to 14 gain/ loss properties of protein structure and function.<sup>129</sup> The use of this system has been widely validated in the context of mtDNA studies,<sup>124</sup> and performs better in an accuracy test when compared with several other methods.<sup>128</sup> Therefore the question can be asked whether individuals in the disease group are impacted on by a combination of rare (mildly deleterious) variants or simply whether such variants are more common in the disease cohort than in the controls.

The mutational load approach moves away from the study of haplogroups and looks at the collective effect of rare (or recent) variants, which are more likely to be deleterious. It distils the likely impact of a person's mtDNA variation into a single value on a continuous scale rather than a letter. Consequently it will have more statistical power than conventional haplogroupassociation studies, as more powerful parametric statistics can be applied and fewer comparisons are required.<sup>130</sup> It offers an alternative method to explore the involvement of mtDNA variants in disease phenotypes, including diseases thought to be related to mitochondrial dysfunction, such as CVDs.

# The unique challenges faced by studies in African populations

While communicable diseases are still the leading causes of mortality in sub-Saharan Africa (SSA), CVD particularly is a growing concern here, since the prevalence has risen markedly in recent times as more populations of developing countries become urbanised and are exposed to a diet and lifestyle that increase risk factors for CVD.<sup>131</sup> Taking into account the many differences among ethnic groups in the onset and development of CVD,<sup>132,133</sup> genomic investigations have also been used to investigate these disparities.<sup>131,134,136</sup> However, the number of well-powered genetic studies on CVDs in African populations or people of African descent is much lower than in European populations. As yet, no conclusive nDNA genetic factor/s has been identified to help understand these disparities.<sup>137</sup>

Current Eurocentric reference panels used in GWAS studies to examine the involvement of population variants in disease have been shown to be of limited use in even common SSA population groups.<sup>138</sup> This is indicative of the lack of African representation in our current databases. This lack extends to mtDNA as well. Of the more than 30 000 mtDNA sequences available on GenBank, only 13% of these are of African lineages (L0–6). This bias in published data results in the resolution of the phylogenetic tree being much higher in the European branches (especially super-haplogroup N descendant) than in the African roots,<sup>139</sup> despite greater diversity within the latter. Comparatively few studies have been done where the involvement of mtDNA variation in CVD has been considered.<sup>135,136,140-142</sup> Although of small size, one such study helps to highlight the challenges posed by these gaps in our current data. Ameh *et al.*<sup>142</sup> could not find the tRNA mutation m.3243A>G in Nigerian type 2 diabetes patients, despite an association being previously reported in other European and Asian populations. This and other studies<sup>143</sup> illustrate the difficulty of extrapolating genetic risk factors for disease from one population group to the next, and the need for population-specific studies.

#### Conclusion

SSA is facing a growing burden of CVD, while the discrepancies in onset and progression between different ethnicities are still poorly understood. Additionally, there are large data gaps when genetic studies on Africans are considered, especially for complex disease phenotypes. The unique genetic backgrounds of different populations also make it difficult to apply advances made in well-studied populations to understudied populations.

While great efforts are being made to address these data gaps by initiatives such as the Human Heredity and Health in Africa (H3Africa) initiative,<sup>131</sup> the Southern African Human Genome Programme, and the African Genome Variation Project,<sup>138</sup> there is an urgent need for even more large-scale African-specific investigations (which should also consider mtDNA variation) to be undertaken if we are to provide the necessary care to all vulnerable groups.<sup>144</sup>

Realistically, for some time still, it is likely that studies in African populations will be hampered by financial and logistic/infrastructural difficulties,<sup>144</sup> limiting the sizes thereof. Fortunately, these studies can benefit from retrospective lessons we have learned thus far in other populations, highlighted in the above discussions. New studies could particularly benefit by asking better-formulated questions, and using alternative approaches that aim to address the challenges associated with many of the classic approaches used, when the role of mtDNA in common disease is investigated.

#### References

- Mensah GA. Descriptive epidemiology of cardiovascular risk factors and diabetes in Africa. *Prog Cardiovasc Dis* 2013; 56: 240–250. http:// dx.doi.org/10.1016/j.pcad.2013.10.014.
- McPherson R, Pertsemlidis A, Kavaslar N, Stewart A, Roberts R, Cox DR, et al. A common allele on chromosome 9 associated with coronary heart disease. Science 2007; 316(5830): 1488–1491. doi:10.1126/ science.1142447.
- Matarín M, Brown W, Scholz S, Simón-Sánchez J, Fung H-C, Hernandez D, et al. A genome-wide genotyping study in patients with ischaemic stroke: initial analysis and data release. *Lancet Neurol* 2007; 6: 414–420. doi:10.1016/s1474-4422(07)70081-9.
- Den Hoed M, Strawbridge RJ, Almgren P, Gustafsson S, Axelsson T, Engström G, et al. GWAS-identified loci for coronary heart disease are associated with intima-media thickness and plaque presence at the carotid artery bulb. *Atherosclerosis* 2015; 239: 304–310. http://dx.doi. org/10.1016/j.atherosclerosis.2015.01.032.
- Chen X, Kuja-Halkola R, Rahman I, Arpegärd J, Viktorin A, Karlsson R, et al. Dominant genetic variation and missing heritability for human complex traits: insights from twin versus genome-wide common SNP models. Am J Hum Genet 2015; 97: 708–714. http://dx.doi.org/10.1016/j. ajhg.2015.10.004.

- Arking DE, Chakravarti A. Understanding cardiovascular disease through the lens of genome-wide association studies. *Trends Genet* 2009; 25(9): 387–394. doi:10.1016/j.tig.2009.07.007.
- Liu C, Yang Q, Hwang S, Sun F, Johnson AD, Shirihai OS, et al. Association of genetic variation in the mitochondrial genome with blood pressure and metabolic traits. *Hypertension* 2012; 60: 949–956. doi: 10.1161/hypertensionaha.112.196519.
- Lotta L. Genome-wide association studies in atherothrombosis. *Eur J* Intern Med 2010; 21: 74–78. doi:10.1016/j.ejim.2009.11.003.
- Peden J, Farrall M. Thirty-five common variants for coronary artery disease: the fruits of much collaborative labour. *Hum Mol Genet* 2011; 20(2): R198–R205. doi:10.1093/hmg/ddr384.
- The CARDIoGRAMplusC4D Consortium. Large-scale association analysis identifies new risk loci for coronary artery disease. *Nat Genet* 2013; 45(1): 25–33. doi: 10.1038/ng.2480.
- Smith JG, Newton-Cheh C. Genome-wide association studies of lateonset cardiovascular disease. J Molec Cell Cardiol 2015; 83: 131–141. http://dx.doi.org/10.1016/j.yjmcc.2015.04.004.
- Andrews RM, Kubacka I, Chinnery PF, Lightowlers RN, Turnbull DM, Howell N. Reanalysis and revision of the Cambridge reference sequence for human mitochondrial DNA. *Nat Genet* 1999; 23: 147. doi:10.1038/13779.
- Marín-García J, Akhmedov A. Mitochondrial dynamics and cell death in heart failure. *Heart Fail Rev* 2016; 21(2):123–136. doi: 10.1007/ s10741-016-9530-2.
- Tuppen HA, Blakely EL, Turnbull DM, Taylor RW. Mitochondrial DNA mutations and human disease. *Biochim Biophys Acta* 2010; 1797(2): 113–128. doi: 10.1016/j.bbabio.2009.09.005.
- de Champlain J, Wu R, Girouard H, Karas M, EL Midaoui A, Laplante M, Wu L. Oxidative stress in hypertension. *Clin Exp Hypertens* 2004; 26(7–8): 593–601. PMID: 15702613.
- Salim S, Asghar M, Chugh G, Taneja M, Xia Z, Saha K. Oxidative stress: A potential recipe for anxiety hypertension and insulin resistance. *Brain Res* 2010; **359**: 178–185. doi:10.1016/j.brainres.2010.08.093.
- Queisser N, Schupp N. Aldosterone oxidative stress and NF-kB activation in hypertension-related cardiovascular and renal diseases. *Free Radic Biol Med* 2012; 53: 314–327. http://dx.doi.org/10.1016/j.freerad-biomed.2012.05.011.
- López-Armada MJ, Riveiro-Naveira RR, Vaamonde-García C, Valcárcel-Ares MN. Mitochondrial dysfunction and the inflammatory response. *Mitochondrion* 2013; 13: 106–118. http://dx.doi.org/10.1016/j. mito.2013.01.003.
- Nakayama H, Otsu K. Translation of hemodynamic stress to sterile inflammation in the heart. *Trends Endocrin Metab* 2013; 24(11): 546–553. http://dx.doi.org/10.1016/j.tem.2013.06.004.
- Van der Walt C, Malan L, Uys AS, Malan NT. Low grade inflammation and ECG left ventricular hypertrophy in urban African males: The SABPA study. *Heart Lung Circ* 2013; **22**(11): 924–929. doi: 10.1016/j. hlc.2013.03.075.
- Harrison DG, Gongora MC, Guzik TJ, Widder J. Oxidative stress and hypertension. J Am Soc Hypertens 2007; 1(1): 30–44. doi:10.1016/j. jash.2006.11.006.
- Gönenç A, Hacışevk A, Tavil Y, Çengel A, Torun M. Oxidative stress in patients with essential hypertension: A comparison of dippers and nondippers. *Eur J Intern Med* 2013; 24: 139–144. http://dx.doi.org/10.1016/j. ejim.2012.08.016.
- Yu EP, Bennett MR. The role of mitochondrial DNA damage in the development of atherosclerosis. *Free Radic Biol Med* 2016; 100: 223–230. doi: 10.1016/j.freeradbiomed.2016.06.011.
- 24. Dantas AP, Franco M, d'Silva-Antonialli MM, Tostes RC, Fortes ZB,

#### Key messages

- Cardiovascular disease (CVD) is a leading global cause of morbidity and mortality, and its incidence is on the rise in sub-Saharan Africa.
- Discrepancies in the onset and progression of CVDs exist between different ethnic and population groups, which nuclear genetic studies have so far failed to explain. Mitochondrial DNA (mtDNA) offers a viable alternative target for genetic studies concerning common complex disease.
- Many approaches can be taken to investigate the role of mtDNA in disease, but not all are suited for studies influenced by moderate cohort size or population stratification. The adjusted mutational load hypothesis offers an alternative approach, which could be of particular value for much-needed studies on CVDs in under-represented sub-Saharan African populations.

Nigro D, *et al.* Gender differences in superoxide generation in microvessels of hypertensive rats: role of NAD(P)H-oxidase. *Cardiovasc Res* 2004; **61**: 22–29. doi:10.1016/j.cardiores.2003.10.010.

- Brière J, Chrétien D, Bénit P, Rustin P. Respiratory chain defects: what do we know for sure about their consequences in vivo? *Biochim Biophys Acta Bioenergetics* 2004; 1659(2): 172–177. http://dx.doi.org/10.1016/j. bbabio.2004.07.002.
- Reinecke F, Smeitink J, Van der Westhuizen FH. OXPHOS gene expression and control in mitochondrial disorders. *Biochim Biophys Acta Molec Basis Dis* 2009; **1792**(12): 1113–1121. doi: 10.1016/j. bbadis.2009.04.003.
- Naviaux R. Metabolic features of the cell danger response. *Mitochondrion* 2014; 16: 7–7. doi: 10.1016/j.mito.2013.08.006.
- Stiefel P, Argüelles S, García S, Jiménez L, Aparicio R, Carneado J, *et al.* Effects of short-term supplementation with folic acid on different oxidative stress parameters in patients with hypertension. *Biochim Biophys Acta* 2005; **1726**: 152–159. doi:10.1016/j.bbagen.2005.07.014.
- Yamaguchi Y, Yamada K, Yoshikawa N, Nakamura K, Haginaka J, Kunitomo M. Corosolic acid prevents oxidative stress inflammation and hypertension in SHR/NDmcr-cp rats a model of metabolic syndrome. *Life Sci* 2006; **79**: 2474–2479. doi:10.1016/j.lfs.2006.08.007.
- Collins LV, Hajizadeh S, Holme E, Jonsson IM, Tarkowski A. Endogenously oxidized mitochondrial DNA induces *in vivo* and *in vitro* inflammatory responses. *J Leukoc Biol* 2004; 75: 995–1000.
- Zhou R, Yazdi A, Menu P, Tschopp J. A role for mitochondria in NLRP3 inflammasome activation. *Nature* 2011; 469(7329): 221–225. doi: 10.1038/nature09663.
- Nakahira K, Haspel J, Rathinam V, Lee S, Dolinay T, Lam H, *et al.* Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat Immunol* 2011; **12**(3): 222–230. doi: 10.1038/ni.1980.
- Oka T, Hikoso S, Yamaguchi O, Taneike M, Takeda T, Tamai Tet al. Mitochondrial DNA that escapes from autophagy causes inflammation and heart failure. *Nature* 2012; 485(7397): 251–255. doi: 10.1038/ nature10992.
- Shimada K, Crother T, Karlin J, Dagvadorj J, Chiba N, Chen S, *et al.* Oxidized mitochondrial DNA activates the NLRP3 inflammasome during apoptosis. *Immunity* 2012; 36(3): 401–414. doi: 10.1016/j. immuni.2012.01.009.
- 35. West A, Khoury-Hanold W, Staron M, Tal M, Pineda C, Lang S, et al.

Mitochondrial DNA stress primes the antiviral innate immune response. *Nature* 2015; **520**(7548): 553–537. doi:10.1038/nature14156.

- 36. Yu E, Calvert P, Mercer J, Harrison J, Baker L, Figg N, et al. Mitochondrial DNA damage can promote atherosclerosis independently of reactive oxygen species through effects on smooth muscle cells and monocytes and correlates with higher-risk plaques in humans. *Circulation* 2013; **128**: 702–712. doi: 10.1161/circulationaha.113.002271.
- Mercer JR. Mitochondrial bioenergetics and therapeutic intervention in cardiovascular disease. *Pharmacol Therapeut* 2014; 141(1):13–20. http:// dx.doi.org/10.1016/j.pharmthera.2013.07.011.
- Yu EP, Bennett MR. Mitochondrial DNA damage and atherosclerosis. *Trends Endocrin Metab* 2014; 25(9): 481–487. http://dx.doi. org/10.1016/j.tem.2014.06.008.
- Lightowlers RN, Taylor RW, Turnbull DM. Mutations causing mitochondrial disease: What is new and what challenges remain? *Science* 2015; **349**(6255): 1494–1499. doi: 10.1126/science.aac7516.
- DiMauro S, Schon E. Mitochondrial DNA mutations in human disease. *Am J Med Genet* 2001; **106**:18–26. doi: 10.1002/ajmg.1392.
- McFarland R, Elson JL, Taylor RW, Howell N, Turnbull DM. Assigning pathogenicity to mitochondrial tRNA mutations: when 'definitely maybe' is not good enough. *Trends Genet* 2004; 20: 591–596. doi: 10.1016/j.tig.2004.09.014.
- Montoya J, López-Gallardo E, Díez-Sánchez C, López-Pérez MJ, Ruiz-Pesini E. 20 years of human mtDNA pathologic point mutations: Carefully reading the pathogenicity criteria. *Biochim Biophys Acta* 2009; 1787: 476–483. doi:10.1016/j.bbabio.2008.09.003.
- Yarham JW, Al-Dosary M, Blakely EL, Alston CL, Taylor RW, Elson JL, *et al.* A comparative analysis approach to determining the pathogenicity of mitochondrial tRNA mutations. *Hum Mutat* 2011; 32(11): 1319–1325. doi: 10.1002/humu.21575.
- Mitchell AL, Elson JL, Howell N, Taylor RW, Turnbull DM. Sequence variation in mitochondrial complex I genes: mutation or polymorphism? *J Med Genet* 2006; 43: 175–179. doi: 10.1136/jmg.2005.032474.
- Rossignol R, Faustin B, Rocher C, Malgat M, Mazat J, Letellier T. Mitochondrial threshold effects. *Biochem J* 2003; **370**(Pt 3): 751–762. doi: 10.1042/bj20021594.
- Faustin B, Rossignol R, Rocher C, Bénard G, Malgat M, Letellier T. Mobilization of adenine nucleotide translocators as molecular bases of the biochemical threshold effect observed in mitochondrial diseases. J Biol Chem 2004; 279(19): 20411–20421. doi 10.1074/jbc.m314259200.
- Picard M, Hirano M. Disentangling (epi)genetic and environmental contributions to the mitochondrial 3243A>G mutation phenotype: Phenotypic destiny in mitochondrial disease? J Am Med Assoc Neurol 2016; 73(8): 923–925. doi: 10.1001/jamaneurol.2016.1676.
- Kirches E. LHON: Mitochondrial mutations and more. *Curr Genomics* 2011; **12**: 44–54. doi: 10.2174/138920211794520150.
- Kirkman M, Yu-Wai-Man P, Korsten A, Leonhardt M, Dimitriadis K, De Coo I, *et al.* Gene–environment interactions in Leber hereditary optic neuropathy. *Brain* 2009; **132**(Pt 9): 2317–2326. doi: 10.1093/brain/ awp158.
- Hudson G, Carelli V, Spruijt L, Gerards M, Mowbray C, Achilli A, *et al.* Clinical expression of Leber hereditary optic neuropathy is affected by the mitochondrial DNA-haplogroup background. *Am J Hum Genet* 2007; **81**(2): 228–233. http://dx.doi.org/10.1086/519394.
- Ji Y, Zhang A, Jia X, Zhang Y, Xiao X, Li S, *et al*. Mitochondrial DNA haplogroups M7b1'2 and M8a affect clinical expression of Leber hereditary optic neuropathy in Chinese families with m.11778G>A mutation. *Am J Hum Genet* 2008; 83(6): 760–768. doi: 10.1016/j.ajhg.2008.11.002.
- 52. Ong S-B, Kalkhoran S, Cabrera-Fuentes H, Hausenloy D. Mitochondrial fusion and fission proteins as novel therapeutic targets for treating cardi-

ovascular disease. Eur J Pharmacol 2015; **763**:104–114. http://dx.doi. org/10.1016/j.ejphar.2015.04.056.

- Finsterer J, Kothari S. Cardiac manifestations of primary mitochondrial disorders. *Int J Cardiol* 2014; 177: 754–763. http://dx.doi.org/10.1016/j. ijcard.2014.11.014.
- Yaplito-Lee J, Weintraub R, Jamsen K, Chow C, Thorburn D, Boneh A. Cardiac manifestations in oxidative phosphorylation disorders of childhood. *J Pediatr* 2007; **150**(4): 407–411. doi: http://dx.doi.org/10.1016/j. jpeds.2006.12.047.
- Limongelli G, Masarone D, D'Alessandro R, Elliott PM. Mitochondrial diseases and the heart: an overview of molecular basis, diagnosis, treatment and clinical course. *Future Cardiol* 2012; 8(1):71–88. doi: 10.2217/ fca.11.79.
- Villar P, Bretón B, García-Pavía P, González-Páramos C, Blázquez A, Gómez-Bueno M, *et al.* Cardiac dysfunction in mitochondrial disease. *Circ J* 2013; 77(11): 2799–2806. doi: 10.1253/circj.CJ-13-0557.
- Yajima N, Yazaki Y, Yoshida K, Sano K, Takahashi W, Sasaki Y, *et al.* A case of mitochondrial cardiomyopathy with pericardial effusion evaluated by 99mTc-MIBI myocardial scintigraphy. *J Nucl Cardiol* 2009; 16(6): 989–994. doi:10.1007/s12350-009-9149-y.
- Malfatti E, Laforêt P, Jardel C, Stojkovic T, Behin A, Eymard B, *et al.* High risk of severe cardiac adverse events in patients with mitochondrial m. 3243A> G mutation. *Neurology* 2013; 80(1): 100–105. doi: http://dx. doi.org/10.1212/WNL.0b013e31827b1a2f.
- Finsterer J, Stöllberger C, Kopsa W. Noncompaction on cardiac MRI in a patient with nail-patella syndrome and mitochondriopathy. *Cardiology* 2003; 100(1): 48–49. doi:10.1159/000072393.
- Inamori M, Ishigami T, Takahashi N, Hibi K, Ashino K, Sumita S, et al. A case of mitochondrial cardiomyopathy with heart failure, sick sinus syndrome and diabetes mellitus: mitochondrial DNA adenine-toguanine transition at 3243 of mitochondrial tRNA (LEU)(UUR) gene. J Cardiol 1997; 30(6): 341–347. PMID: 9436076.
- Majamaa-Voltti K, Peuhkurinen K, Kortelainen ML, Hassinen IE, Majamaa K. Cardiac abnormalities in patients with mitochondrial DNA mutation 3243A> G. *BMC Cardiovasc Disord* 2002; 2(1): 1. doi: 10.1186/1471-2261-2-12.
- Hollingsworth KG, Gorman GS, Trenell MI, McFarland R, Taylor RW, Turnbull DM, *et al.* Cardiomyopathy is common in patients with the mitochondrial DNA m. 3243A> G mutation and correlates with mutation load. *Neuromusc Disord* 2012; 22(7): 592–596. doi: http://dx.doi. org/10.1016/j.nmd.2012.03.001.
- Mima A, Shiota F, Matsubara T, Iehara N, Akagi T, Abe H, *et al*. An autopsy case of mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes (MELAS) with intestinal bleeding in chronic renal failure. *Renal Fail* 2011; 33(6): 622–625. http://dx.doi.org/10.3109/0886022X.2011.585730.
- Vydt TC, de Coo RF, Soliman OI, Folkert J, van Geuns RJ, Vletter WB, et al. Cardiac involvement in adults with m. 3243A> G MELAS gene mutation. Am J Cardiol 2007; 99(2): 264–269. http://dx.doi. org/10.1016/j.amjcard.2006.07.089.
- Hung PC, Wang HS, Chung HT, Hwang MS, Ro LS. Pulmonary hypertension in a child with mitochondrial A3243G point mutation. *Brain Develop* 2012; 34(10): 866–868. http://dx.doi.org/10.1016/j.braindev.2012.02.011.
- Liu CH, Chang CH, Kuo HC, Ro LS, Liou CW, Wei YH, et al. Prognosis of symptomatic patients with the A3243G mutation of mitochondrial DNA. J Formosan Med Assoc 2012; 111(9): 489–494. http:// dx.doi.org/10.1016/j.jfma.2011.06.014.
- 67. Sproule DM, Dyme J, Coku J, de Vinck D, Rosenzweig E, Chung WK, *et al.* Pulmonary artery hypertension in a child with MELAS due to

a point mutation of the mitochondrial tRNA (Leu) gene (m. 3243A>G). *J Inherit Metab Dis* 2008; **31**(3): 497–503. doi:10.1007/s10545-007-0735-3.

- Wang W, Seak CJ, Liao SC, Chiu TF, Chen JC. Cardiac tamponade: a new complication in a patient with mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke like episodes. *Am J Emerg Med* 2008; 26(3): 382–e1. doi: 10.1016/j.ajem.2007.05.027.
- Majamaa-Voltti K, Majamaa K, Peuhkurinen K, Mäkikallio T, Huikuri H. Cardiovascular autonomic regulation in patients with 3243A> G mitochondrial DNA mutation. *Ann Med* 2004; 36(3): 225–231. http:// dx.doi.org/10.1080/07853890410028456.
- Sweeney MG, Brockington M, Weston MJ, Morgan-Hughes JA, Harding AE. Mitochondrial DNA transfer RNA mutation Leu (UUR) A→ G 3260: a second family with myopathy and cardiomyopathy. Q J Med 1993; 86(7): 435–438. doi: http://dx.doi.org/ 435-438. PMID: 8210299.
- Zeviani M, Gellera C, Antozzi C, Rimoldi M, Morandi L, Tiranti V, et al. Maternally inherited myopathy and cardiomyopathy: association with mutation in mitochondrial DNA tRNALeu (UUR). *Lancet* 1991; 338(8760): 143–147. doi:10.1016/0140-6736(91)90136-D.
- Barišic N, Kleiner IM, Malcic I, Papa J, Boranic M. Spinal dysraphism associated with congenital heart disorder in a girl with MELAS syndrome and point mutation at mitochondrial DNA nucleotide 3271. *Croat Med J* 2002; 43(1): 37–41. PMID: 11828557.
- Silvestri G, Santorelli FM, Shanske S, Whitley CB, Schimmenti LA, Smith SA, *et al.* A new mtDNA mutation in the tRNALeu (UUR) gene associated with maternally inherited cardiomyopathy. *Hum Mutat* 1994; 3(1): 37–43. doi: 10.1002/humu.1380030107.
- Palecek T, Tesarova M, Kuchynka P, Dytrych V, Elleder M, Hulkova H, *et al.* Hypertrophic cardiomyopathy due to the mitochondrial DNA mutation m. 3303C> T diagnosed in an adult male. *Int Heart J* 2012; 53(6): 383–387. http://doi.org/10.1536/ihj.53.383.
- 75. Hayashi J, Ohta S, Kagawa Y, Takai D, Miyabayashi S, Tada K, et al. Functional and morphological abnormalities of mitochondria in human cells containing mitochondrial DNA with pathogenic point mutations in tRNA genes. J Biol Chem 1994; 269(29): 19060–19066. PMID: 7518448.
- Taniike M, Fukushima H, Yanagihara I, Tsukamoto H, Tanaka J, Fujimura H, *et al.* Mitochondrial tRNAlle mutation in fatal cardiomyopathy. *Biochem Biophys Res Co* 1992; 186(1): 47–53. doi:10.1016/ s0006-291x(05)80773-9.
- Giordano C, Perli E, Orlandi M, Pisano A, Tuppen HA, He L, et al. Cardiomyopathies due to homoplasmic mitochondrial tRNA mutations: morphologic and molecular features. *Hum Pathol* 2013; 44(7): 1262–1270. http://dx.doi.org/10.1016/j.humpath.2012.10.011.
- Casali C, Santorelli FM, Damati G, Bernucci P, DeBiase L, DiMauro S. A novel mtDNA point mutation in maternally inherited cardiomyopathy. *Biochem Biophys Res Co* 1995; 213(2): 588–593. doi:10.1006/ bbrc.1995.2172.
- Sacconi S, Salviati L, Nishigaki Y, Walker WF, Hernandez-Rosa E, Trevisson E, et al. A functionally dominant mitochondrial DNA mutation. *Hum Mol Genet* 2008; 17(12): 1814–1820. doi: 10.1093/hmg/ ddn073.
- Villar P, Bretón B, García-Pavía P, González-Páramos C, Blázquez A, Gómez-Bueno M, *et al.* Cardiac dysfunction in mitochondrial disease. *Circ J* 2013; 77(11): 2799–2806. http://doi.org/10.1253/circj.CJ-13-0557.
- Vallance HD, Jeven G, Wallace DC, Brown MD. A case of sporadic infantile histiocytoid cardiomyopathy caused by the A8344G (MERRF) mitochondrial DNA mutation. *Pediatr Cardiol* 2004; 25(5): 538–540. doi:10.1007/s00246-003-0446-y.
- 82. Wahbi K, Larue S, Jardel C, Meune C, Stojkovic T, Ziegler F, et al.

Cardiac involvement is frequent in patients with the m. 8344A>G mutation of mitochondrial DNA. *Neurology* 2010; **74**(8): 674–677. http://dx. doi.org/10.1212/WNL.0b013e3181d0ccf4.

- Santorelli FM, Mak SC, El-Schahawi M, Casali C, Shanske S, Baram TZ, et al. Maternally inherited cardiomyopathy and hearing loss associated with a novel mutation in the mitochondrial tRNA (Lys) gene (G8363A). Am J Hum Genet 1996; 58(5): 933. PMCID: PMC1914622.
- Jonckheere AI, Hogeveen M, Nijtmans LG, van den Brand MA, Janssen AJ, Diepstra JH, *et al*. A novel mitochondrial ATP8 gene mutation in a patient with apical hypertrophic cardiomyopathy and neuropathy. *J Med Genet* 2008; 45(3): 129–133. doi:10.1136/jmg.2007.052084.
- Chamkha I, Alila-Fersi O, Mkaouar-Rebai E, Aloulou H, Kifagi C, Hachicha M, et al. A novel m. 12908T> A mutation in the mitochondrial ND5 gene in patient with infantile-onset Pompe disease. Biochem Biophys Res Co 2012; 429(1): 31–38. http://dx.doi.org/10.1016/j. bbrc.2012.10.105.
- Wang SB, Weng WC, Lee NC, Hwu WL, Fan PC, Lee WT. Mutation of mitochondrial DNA G13513A presenting with Leigh syndrome, Wolff– Parkinson–White syndrome and cardiomyopathy. *Pediat Neonatol* 2008; 49(4): 145–149. doi:10.1016/S1875-9572(08)60030-3.
- Van Hove JL, Freehauf C, Miyamoto S, Vladutiu GD, Pancrudo J, Bonilla E, *et al.* Infantile cardiomyopathy caused by the T14709C mutation in the mitochondrial tRNA glutamic acid gene. *Eur J Pediatr* 2008; 167(7): 771–776. doi:10.1007/s00431-007-0587-8.
- Andreu AL, Checcarelli N, Iwata S, Shanske S, Dimauro S. A missense mutation in the mitochondrial cytochrome b gene in a revisited case with histiocytoid cardiomyopathy. *Pediatr Res* 2000; 48(3): 311–314. doi:10.1203/00006450-200009000-00008.
- Sahin E, DePinho R. Axis of ageing: telomeres p53 and mitochondria. *Nat Rev Molec Cell Biol* 2012; 13(6): 397–404. doi: 10.1038/nrm3352.
- Laderman K, Penny J, Mazzucchelli F, Bresolin N, Scarlato G, Attardi G. Aging-dependent functional alterations of mitochondrial DNA mtDNA from human fibroblasts transferred into mtDNA-less cells. J Biol Chem 1996; 271: 15891–15897. PMID: 8663253.
- Pyle A, Anugrha H, Kurzawa-Akanbi M, Yarnall A, Burn D, Hudson G. Reduced mitochondrial DNA copy number is a biomarker of Parkinson's disease. *Neurobiol Aging* 2016; 38: 216.e7-216.e10. http:// dx.doi.org/10.1016/j.neurobiolaging.2015.10.033.
- Gui Y-X, Xu Z-P, Lv W, Zhao J-J, Hu X-Y. Evidence for polymerase gamma POLG1 variation in reduced mitochondrial DNA copy number in Parkinson's disease. *Parkinsonism Related Disord* 2015; 21: 282–286. http://dx.doi.org/10.1016/j.parkreldis.2014.12.030.
- Malik AN, Parsade CK, Ajaz S, Crosby-Nwaobi R, Gnudi L, Czajka A, et al. Altered circulating mitochondrial DNA and increased inflammation in patients with diabetic retinopathy. *Diabetes Res Clin Pract* 2015; 110(3): 257–265. doi: 10.1016/j.diabres.2015.10.006.
- Czajka A, Ajaz S, Gnudi L, Parsade CK, Jones P, Reid F, et al. Altered mitochondrial function, mitochondrial DNA and reduced metabolic flexibility in patients with diabetic nephropathy. *EBioMedicine* 2015; 2(6): 499–512. doi: http://dx.doi.org/10.1016/j.ebiom.2015.04.002.
- Tyrka AR, Carpenter LL, Kao H-T, Porton B, Philipa NS, Ridout SJ, et al. Association of telomere length and mitochondrial DNA copy number in a community sample of healthy adults. *Exp Gerontol* 2015; 66: 17–20. http://dx.doi.org/10.1016/j.exger.2015.04.002.
- Lagouge M, Larsson N. The role of mitochondrial DNA mutations and free radicals in disease and ageing. *J Intern Med* 2013; 273(6): 529–543. doi: 10.1111/joim.12055.
- 97. Pisano A, Cerbelli B, Perli E, Pelullo M, Bargelli V, Preziuso C, et al. Impaired mitochondrial biogenesis is a common feature to myocardial hypertrophy and end-stage ischemic heart failure. *Cardiovasc Pathol*

2016; 25(2): 103-112. http://dx.doi.org/10.1016/j.carpath.2015.09.009.

- Hao X-D, Chen P, Wang Y, Li S-X, Xie L-X. Mitochondrial DNA copy number but not haplogroup is associated with keratoconus in Han Chinese population. *Exp Eye Res* 2015; **132**: 59–63. http://dx.doi. org/10.1016/j.exer.2015.01.016.
- Bersani FS, Morley C, Lindqvist D, Epel ES, Picard M, Yehuda R, *et al.* Mitochondrial DNA copy number is reduced in male combat veterans with PTSD. *Prog Neuro-Psychoph* 2016; 64: 10–17. http://dx.doi. org/10.1016/j.pnpbp.2015.06.012.
- Van Oven M, Kayser M. Updated comprehensive phylogenetic tree of global human mitochondrial DNA variation. *Hum Mutat* 2009; **30**(2): E386–394. doi: 10.1002/humu.20921.
- 101. Hudson G, Gomez-Duran A, Wilson IJ, Chinnery PF. Recent mitochondrial DNA mutations increase the risk of developing common late-onset human diseases. *PLoS Genet* 2014; **10**(5): e1004369. doi:10.1371/journal. pgen.1004369.
- 102. Strauss KA, DuBiner L, Simon M, Zaragoza M, Sengupta PP, Li P, et al. Severity of cardiomyopathy associated with adenine nucleotide translocator-1 deficiency correlates with mtDNA haplogroup. Proc Natl Acad Sci USA 2013; 110: 3453–3458. doi: 10.1073/pnas.1300690110.
- 103. Mishmar D, Ruiz-Pesini E, Golik P, Macaulay V, Clark AG, Hosseini S, et al. Natural selection shaped regional mtDNA variation in humans. Proc Natl Acad Sci USA. 2003; 100(1): 171–176. doi:10.1073/pnas.0136972100.
- 104. Ji F, Sharpley M, Derbeneva O, Alves L, Qian P, Wang Y, et al. Mitochondrial DNA variant associated with Leber hereditary optic neuropathy and high-altitude Tibetans. Proc Natl Acad Sci USA 2012; 109(19): 7391–7396. doi: 10.1086/519394.
- Wallace DC. Mitochondrial DNA variation in human radiation and disease. *Cell* 2015; 163(1): 33–38. doi:10.1016/j.cell.2015.08.067.
- Moilanen J, Finnilä S, Majamaa K. Lineage-specific selection in human mtDNA: lack of polymorphisms in a segment of MTND5 gene in haplogroup J. *Molec Biol Evol* 2003; **20**: 2132–2142. doi: 10.1093/ molbev/msg230.
- Elson JL, Turnbull DM, Howell N. Comparative genomics and the evolution of human mitochondrial DNA: assessing the effects of selection. *Am J Hum Genet* 2004; **74**: 229–238. doi: 10.1086/381505.
- Amo T, Brand M. Were inefficient mitochondrial haplogroups selected during migrations of modern humans? A test using modular kinetic analysis of coupling in mitochondria from cybrid cell lines. *Biochem J* 2007; 404: 345–351. doi:10.1042/bj20061609.
- 109. Kivisild T, Shen P, Wall DP, Do B, Sung R, Davis K, *et al.* The role of selection in the evolution of human mitochondrial genomes. *Genetics* 2006; **172**: 373–387. doi: 10.1534/genetics.105.043901.
- 110. Howell N, Howell C, Elson JL. Molecular clock debate: Time dependency of molecular rate estimates for mtDNA: this is not the time for wishful thinking. *Heredity* 2008; 107–108. doi:10.1038/hdy.2008.52.
- 111. Elson JL, Herrnstadt C, Preston G, Thal L, Morris CM, Edwardson JA, et al. Does the mitochondrial genome play a role in the etiology of Alzheimer's disease? *Hum Genet* 2006; **119**: 241–254. doi 10.1007/s00439-005-0123-8.
- 112. Maruszak A, Żekanowski C. Mitochondrial dysfunction and Alzheimer's disease. *Prog Neuro-Psychoph* 2011; 35(2): 320–330. doi: 10.1016/j.pnpbp.2010.07.004.
- 113. Crispim D, Canani LH, Gross JL, Tschiedel B, Souto KE, Roisenberg I. The European-specific mitochondrial cluster J/T could confer an increased risk of insulin-resistance and type 2 diabetes: an analysis of the m.4216T > C and m.4917A > G variants. *Ann Hum Genet* 2006; **70**(Pt 4): 488–495. doi: 10.1111/j.1469-1809.2005.00249.x.
- 114. Li S, Besenbacher S, Li Y, Kristiansen K, Grarup N, Albrechtsen A, et

*al.* Variation and association to diabetes in 2000 full mtDNA sequences mined from an exome study in a Danish population. *Eur J Hum Genet* 2014; **22**: 1040–1045. doi:10.1038/ejhg.2013.282.

- 115. Chinnery PF, Mowbray C, Patel S, Elson JL, Sampson M, Hitman G, et al. Mitochondrial DNA haplogroups and type 2 diabetes: a study of 897 cases and 1010 controls. J Med Genet 2007; 44(6): e80. http://dx.doi. org/10.1136/jmg.2007.048876.
- 116. Achilli A, Olivieri A, Pala M, Hooshiar Kashani B, Carossa V, Perego UA, *et al.* Mitochondrial DNA backgrounds might modulate diabetes complications rather than T2DM as a whole. *PloS One* 2011; 6(6): e21029. doi:10.1371/journal.pone.0021029.
- 117. Flaquer A, Baumbach C, Kriebel J, Meitinger T, Peters A, Waldenberger M, et al. Mitochondrial genetic variants identified to be associated with BMI in adults. *PLoS One* 2014; 9(8): e105116. doi:10.1371/journal. pone.0105116.
- Chinnery PF, Elliott HR, Syed A, Rothwell PM, Oxford vascular study. Mitochondrial DNA haplogroups and risk of transient ischaemic attack and ischaemic stroke: a genetic association study. *Lancet Neurol* 2010; 9(5): 498–503. doi: 10.1016/S1474-4422(10)70083-1.
- 119. Elango S, Govindaraj P, Vishwanadha VP, Reddy G, Tamang R, Muthusami U, et al. Analysis of mitochondrial genome revealed a rare 50 bp deletion and substitutions in a family with hypertension. *Mitochondrion* 2011; 11: 878–885. doi:10.1016/j.mito.2011.07.002.
- Govindaraj P, Khanb NA, Rani B, Rani DS, Selvaraj P, Jyothi V, *et al.* Mitochondrial DNA variations associated with hypertrophic cardiomyopathy. *Mitochondrion* 2014; 16:65–72. http://dx.doi.org/10.1016/j. mito.2013.10.006.
- 121. Salas A, Elson JL. Raising doubts about the pathogenicity of mitochondrial DNA mutation m.3308T>C in left ventricular hypertraveculation/compactation. *Cardiology* 2011; **122**(2): 113–115. doi: 10.1159/000339348.
- 122. Salas A, Elson JL. Mitochondrial DNA as a risk factor for false positives in case-control association studies. J Genet Genomics 2015; 42(4): 169–172. http://dx.doi.org/10.1016/j.jgg.2015.03.002.
- 123. Samuels DC, Carothers AD, Horton R, Chinnery PF. The power to detect disease associated with mitochondrial DNA haplogroups. *Am J Hum Genet* 2006; **78**(4): 713–720. http://dx.doi.org/10.1086/502682.
- 124. Pereira L, Soares P, Radivoiac P, Li B, Samuels DC. Comparing phylogeny and the predicted pathogenicity of protein variations reveals equal purifying selection across the global human mtDNA diversity. *Am J Hum Genet* 2011; 88(4): 433–439. doi 10.1016/j.ajhg.2011.03.006.
- 125. Nakagawa Y, Ikegami H, Yamato E, Takekawa K, Fujisawa T, Hamada Y, *et al.* A new mitochondrial DNA mutation associated with non-insulin-dependent diabetes mellitus. *Biochim Biophys Res Commun* 1995; 209: 664–668. doi:10.1006/bbrc.1995.1550.
- 126. López-Gallardo E, Iceta R, Iglesias E, Montoya J, Ruiz-Pesini E. OXPHOS toxicogenomics and Parkinson's disease. *Mutat Res Rev Mutat* 2011; 728(3): 98–106. doi: 10.1016/j.mrrev.2011.06.004.
- 127. López-Gallardo E, Llobet L, Emperador S, Montoya J, Ruiz-Pesini E. Effects of tributyltin chloride on cybrids with or without an ATP synthase pathologic mutation. *Environ Health Perspect* 2016; **124**: 1399–1405. http://dx.doi.org/10.1289/ehp182.
- Thusberg J, Olatubosun A, Vihinen M. Performance of mutation pathogenicity prediction methods on missense variants. *Hum Mutat* 2011; 32(4): 358–368. doi: 10.1002/humu.21445.
- 129. Li B, Krishnan VG, Mort ME, Xin F, Kamati KK, Cooper DN, et al. Automated inference of molecular mechanisms of disease from amino acid substitutions. *Bioinformatics* 2009; 25(21): 2744–2750. doi:10.1093/ bioinformatics/btp528.
- 130. Venter M, Malan L, van Dyk E, Elson JL, van der Westhuizen FH.

Using MutPred derived mtDNA load scores to evaluate mtDNA variation in hypertension and diabetes in a two-population cohort: The SABPA study. *J Genet Genomics* 2017; **44**: 139–149. doi:10.1016/j. jgg.2016.12.003.

- 131. Owolabi MO, Mensah GA, Kimmel PA, Adu D, Ramsay M, Waddy SP, et al. Understanding the rise in cardiovascular diseases in Africa: harmonising H3Africa genomic epidemiological teams and tools. Cardiovas J Afr 2014; 25(3): 134–136. doi: 10.5830/cvja-2014-030.
- 132. Sliwa K, Wilkinson D, Hansen C, Ntyintyane L, Tibazarwa K, Becker A, *et al.* Spectrum of heart disease and risk factors in a black urban population in South Africa the Heart of Soweto Study: a cohort study. *Lancet* 2008; **371**: 915–922. doi: 10.1016/S0140-6736(08)60417-1.
- 133. Okin PM, Kjeldsen SE, Dahlöf B, Devereux RB. Racial differences in incident heart failure during antihypertensive therapy. *Circ Cardiovasc Qualitat Outcomes* 2011; 4: 157–164. doi: 10.1161/circoutcomes.110.960112.
- 134. Lai C-Q, Tucker KL, Choudhry S, Parnell LD, Mattei J, García-Bailo B, *et al.* Population admixture associated with disease prevalence in the Boston Puerto Rican health study. *Hum Genet* 2009; **125**: 199–209. doi 10.1007/s00439-008-0612-7.
- 135. Cardena M, Ribeiro-dos-Santos A, Santos S, Mansur A, Pereira A, Fridman C. Amerindian genetic ancestry is associated with higher survival rates compared to African and European ancestry in Brazilian patients with heart failure. *Int J Cardiol* 2014; **176**(2): 527–528. http:// dx.doi.org/10.1016/j.ijcard.2014.07.039.
- 136. Cardena M, Ribeiro-Dos-Santos A, Santos S, Mansur A, Bernardez-Pereira S, Santos P, *et al.* Mitochondrial and genomic ancestry are associated with etiology of heart failure in Brazilian patients. *J Hum Hypertens* 2016; **30**(2): 120–123. doi:10.1038/jhh.2015.39.
- 137. Kaufman JS, Dolman L, Rushani D, Cooper RS. The contribution of genomic research to explaining racial disparities in cardiovascular disease: A systematic review. *Am J Epidemiol* 2015; **181**(7): 464–472. doi: 10.1093/aje/kwu319.

- Gurdasani D, Carstensen T, Tekola-Ayele F, Pagani L, Tachmazidou I, Hatzikotoulas K, *et al.* The African genome variation project shapes medical genetics in Africa. *Nature* 2015; **517**: 327–332. doi:10.1038/nature13997.
- 139. Cavadas B, Soares P, Camacho R, Brandao A, Costa MD, Fernandes V, et al. Fine time scaling of purifying selection on human nonsynonymous mtDNA mutations based on the worldwide population tree and mother–child pairs. *Hum Mutat* 2014; 36(11): 1100–1111. doi: 10.1002/ humu.22849.
- 140. Khogali SS, Myosi BM, Beattie JM, McKenna WJ, Watkins H, Poulton JA. A common mitochondrial DNA variant associated with susceptibility to dilated cardiomyopathy in two different populations. *Lancet* 2001; 357: 1265–1267. doi:10.1016/S0140-67360004422-6.
- 141. Robinson MT, Fischel-Ghodsian N, Fraser HS, Nicholson GD, Grim CM, Wilson DM, *et al.* Genetic influences on the increase in blood pressure with age in normotensice subjects in Barbados. *Ethnic Dis* 2004; 14: 57–63. PMID: 15002924.
- 142. Ameh J, Godwin I, Obi I, Puepet F, Aminu B, Suleiman T. The search for mitochondrial tRNALeu (UUR) A3243G mutation among type 2 diabetes mellitus patients in the Nigerian population. *Afr J Biotechnol* 2011; **10**(62): 13383–13389. doi: 10.5897/ajb11.1556.
- 143. Van der Walt EM, Smuts I, Taylor RW, Elson JL, Turnbull DM, Louw R, et al. Characterization of mtDNA variation in a cohort of South African paediatric patients with mitochondrial disease. J Hum Genet 2012; 20(6): 650–656. doi: 10.1038/ejhg.2011.262.
- 144. Van der Westhuizen FH, Sinxadi PZ, Dandara C, Smuts I, Riordan G, Meldau S, *et al.* Understanding the implications of mitochondrial DNA variation in the health of black southern African populations: The 2014 workshop. *Hum Mutat* 2015; **36**(5): 569–571. doi: 10.1002/humu.22789.
- 145. Forero DA, Wonkam A, Wang W, Laissue P, López-Correa C, Fernández-López JC, et al. Current needs for human and medical genomics research infrastructure in low and middle income countries. J Med Genet 2016; 53(7): 438–440. doi: 10.1136/jmedgenet-2015-103631.